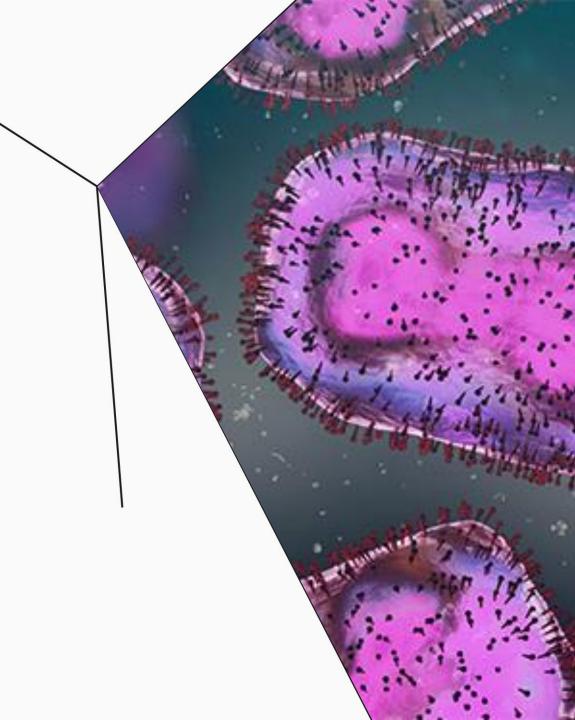
Developing accessible mpox tests

Dr Jennifer Heaney Senior Research Fellow Clinical Immunology Services University of Birmingham





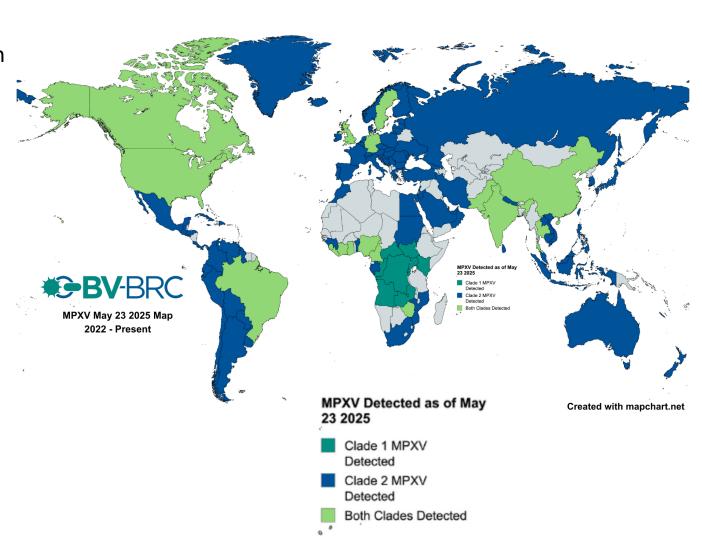
Mpox: a global threat to public health

Different types of Mpox virus (Clade I and Clade II): both have caused WHO Public Health Emergencies of International Concern

Mpox clade II was mainly found in Western Africa until the **2022 outbreak** where it spread worldwide with 102,000 cases. Cases still ongoing......

Mpox clade I endemic in Central Africa and evolution of novel subclade Ib caused **2024 outbreak** to neighbouring countries and other continents

Transmission ongoing in Africa.....



Mpox: diagnostics gap

Evolution of the virus and geographical spread has outpaced development and availability of suitable diagnostic assays

Assay challenges for case detection/acute infection

- 1) case differentiation (Clade I vs II): severity and fatality differences
- 2) Platform and training: centralised reference laboratories
- 3) Lack of rapid diagnostic tests: limited molecular POC tests (PCR) and no suitable rapid antigen (viral protein) POC tests

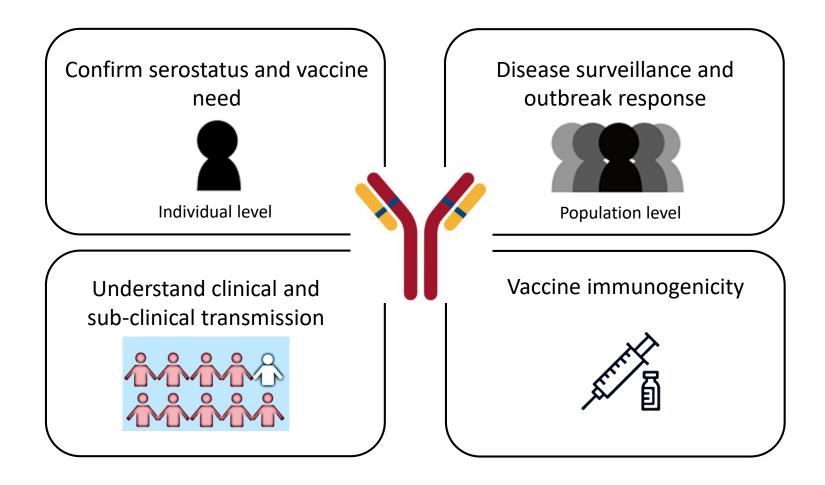


Diagnostics gap global problem



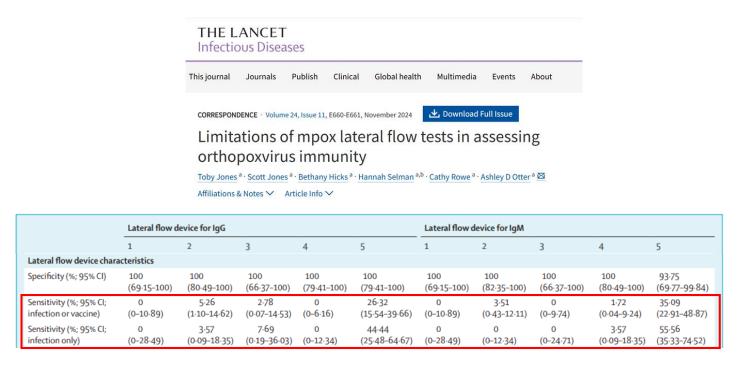
Mpox: immunodiagnostics gap

We have the issue of not only detection of acute infection but surveillance and monitoring, requiring antibody detection



We need affordable and accessible tools that permit antibody detection and quantitation across locations and populations

Immunodiagnostic landscape: PoC tests



Sensitivity ≤ 56% on commercially available lateral flow tests

Problem 1: Lack of point of care immunodiagnostics with acceptable performance characteristics

Immunodiagnostic landscape: lab tests



Luminex multiplex

MpoxPlex



12 antigens



Electrochemiluminescence immunoassay

MSD Orthopoxvirus assay

10 antigens



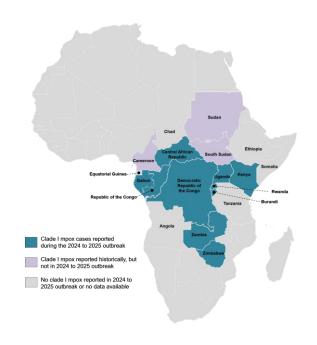
Commercial ELISA kits

Typically single antigen

Problem 2: Real-world application of laboratory testing \rightarrow requires specialised technology platforms and/or the need to measure multiple antigens, with associated cost, time and expertise

Assay translation for resource limited settings







End user needs







Equipment



User friendly



Time

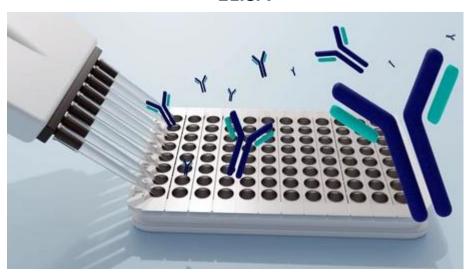


Cost

Assay design for end user needs

1. Available and sustainable platform



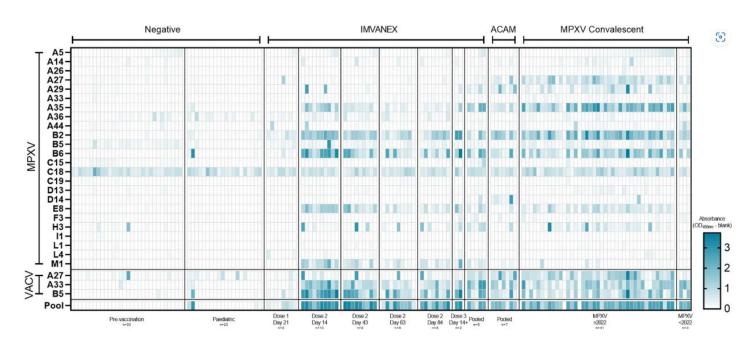


- > Routinely used in clinical laboratories worldwide
- > Allow multiple patient samples to be screened simultaneously
 - > Low cost to manufacture
- > No specialised technology platforms (maintenance, reagent, costs, training)

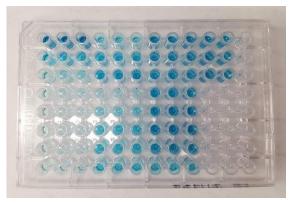
Assay design for end user needs



- 2. Accurately identify and quantify antibodies from infection and/or vaccination
- 3. Lowest number of antigens
- 4. Single readout to minimise complexity and determine serostatus



Combined 4-antigen ELISA



Combination of four immunodominant mpox proteins to enable parallel detection of mpox-specific IgG antibodies derived from both infection and vaccination

Nature Communications volume 14, 5948 (2023)

Assay format for end user needs



Ship to Rwanda or other laboratories

Ease of use within and between laboratories

Simple kit format with recombinant antigen mix dried into plates and pre-dispensed reagents

Development into kit format



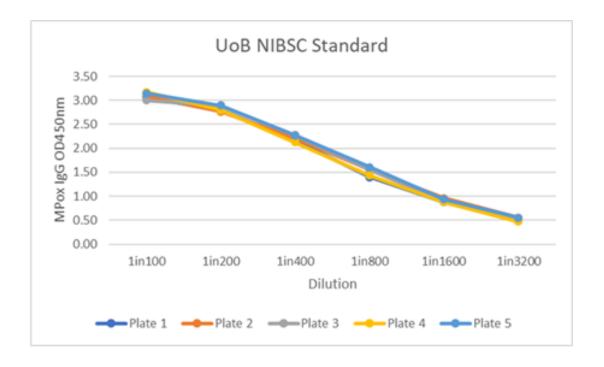


Assay readout for end user needs



There is currently no international reference sera for Mpox with assigned units

However, NIBSC do have a working reagent for anti-mpox antibodies



For each serum sample measured, the assay provides a single data point (OD), which is then adjusted using the NIBSC reagent to provide an IgG antibody ratio

Performance validation UK



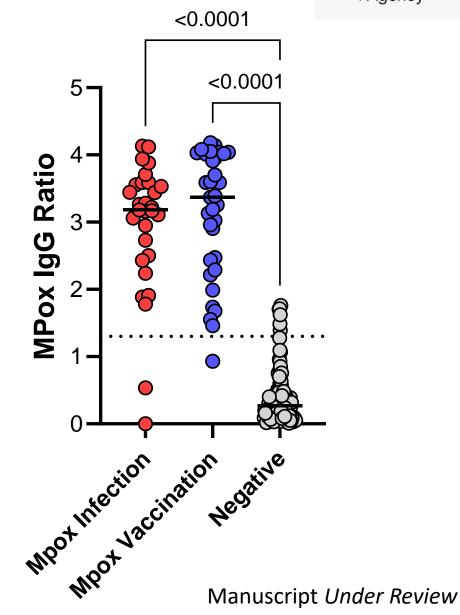


Mpox infection: n = 29 (24-113 days post infection, Clade II)

Mpox vaccinated IMVANEX-vaccinated: (24-77 days post 1st dose [n = 8], 14-84 days post 2nd dose [n = 24], 14 days post 3rd dose [n = 1])

Negative (healthy donors with no known history of mpox infection or IMVANEX vaccination): n = 125

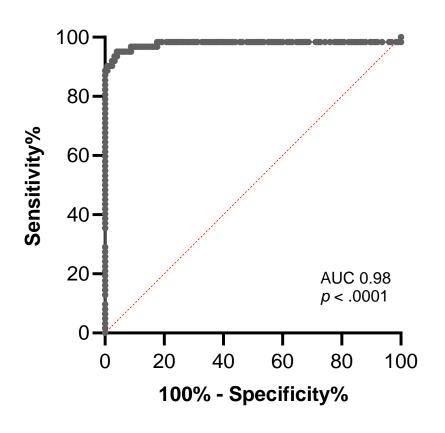
The combined four-antigen ELISA detected significantly higher mpox serum IgG antibodies in both infection and vaccination cohorts compared with negative donors.



Performance validation UK







The ELISA effectively discriminated individuals with prior infection or vaccination from those without

An IgG ratio cut off ≥ 1.33

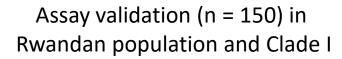
Sensitivity 95% (95% CI 86·71–98·68%)

Specificity 95% (95% CI 89·92–97·78%)

IgG ratio offers an effective quantitative method to distinguish past exposure and/or vaccination, and assess antibody kinetics and change over time

Performance validation Rwanda



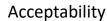


Groups: past infection/vaccinated, controls



Sampling method









Comprehensive Assessment for Responsive Immunisation in Emergency Outbreaks

Groups: past infection/vaccinated, controls, at risk

Aim: Evaluate immunodiagnostics to estimate Mpox's sero-prevalence as a tool to monitor disease burden and prioritise future vaccine utilisation

Immunosurveillance platform established within National Reference Laboratory

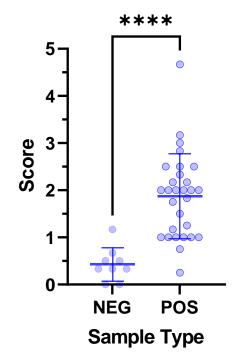


Next steps

- Pending successful validation in Rwanda, explore interest from other laboratories, offer as research tool
- Funding or partnerships to progress (assay optimisation and regulatory approval)
- Address lack of community and PoC antibody tests

Proof of concept for mpox IgG lateral flow test as part of this collaborative project









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UK Health















Training and capacity building

- PhD Scholars
- Laboratory placements







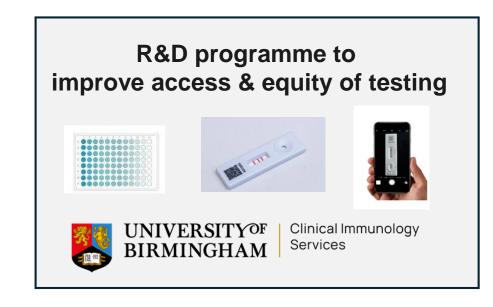
Partner locations

- Rwanda
- South Africa









Evaluation in UK and Africa









Diagnostic areas

Infectious disease and vaccination immunity

Measles, Tetanus

Pandemic preparedness

Mpox, Influenza, H5N1

Screening non-communicable disease

- Autoimmunity T1D and lupus
- Blood cancer myeloma

Partnerships with industry







Project Team



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Dr Matt Pearce Jake Hodgson Rebecca Newman Dr Andrea Murray

Funders





UoB Infrastructure

BUSINESS ENGAGMENT





ENTERPRISE

