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Introduction

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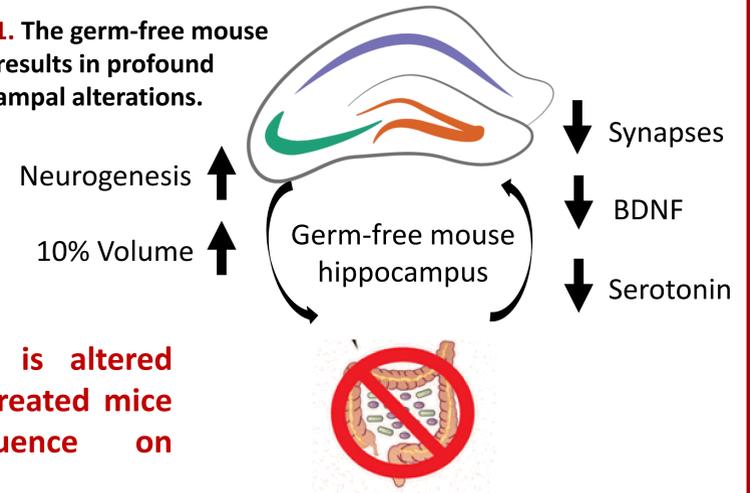
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Previous literature strongly suggests that neuronal circuits and neuronal activity can be influenced by microbiome factors.

- Principally, altered behaviours seen with either supplementation or removal of gut microbiome factors (reviewed in Cryan et al., 2019) demands differential brain activity.
- Germ free mice exhibit altered synaptic plasticity alongside altered behavioural readouts including learning and memory processes (Darch et al., 2021; Gareau et al., 2011; Hoban et al., 2018; Lu et al., 2018; Luk et al., 2018; Pan et al., 2019).
- Morphological differences in germ-free (GF) hippocampus dendrites has been shown (Luczynski et al., 2016).
- Antibiotics decrease hippocampal neurogenesis and memory retention in 6-8 week old mice (Möhle et al., 2016).

Currently, it is unknown whether functional activity of the hippocampus is altered by acute microbiome perturbation in adulthood. Therefore, we used antibiotic-treated mice to determine whether the gut microbiome can exert influence on hippocampal synaptic properties in adulthood.

Figure 1. The germ-free mouse model results in profound hippocampal alterations.



Methods

Multi-electrode array in-vitro hippocampal slice electrophysiology

Transverse hippocampal sections were taken from 3 groups of C57/Bl6 mice aged 10-11wks (antibiotic cocktail-treated (ampicillin 1g/L; vancomycin 0.5g/L and imipenem 0.25g/L) (ABX) (N female/male = 8/9), vancomycin-treated (vancomycin 0.5g/L) (Vanc) (N female/male = 6/9), vehicle control (CTRL) (N female/male = 7/13)), as per standard protocols (Papouin and Haydon, 2018). Antibiotics were delivered in drinking water for 2 weeks. Slices were placed in a multi-electrode array (MEA), stimulating the CA3/CA1 Schaffer collateral pathway. An adjacent electrode, 200um away from the stimulating electrode, recorded field Excitatory Post-Synaptic Potentials (fEPSPs) elicited by the Schaffer collateral axons.

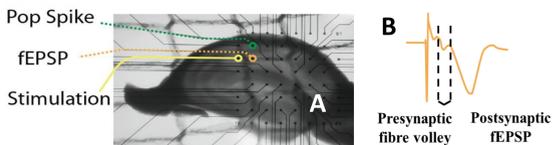


Figure 2. (A). Image of slice with recording and stimulation sites marked. (B). Graph of neuronal population field Excitatory Post Synaptic Potential (fEPSP)

Stimulation protocols:

Input/Output calibration

Slices were stimulated over a range of stimulus intensities between 1-150uA. Subsequent stimuli were calibrated to elicit fEPSP responses at ~40% of the maximum

Paired-Pulse Facilitation

Slices were given double stimuli with inter-pulse intervals ranging between 25 – 200ms. The percent change of the second pulse-response was computed.

Long-Term Potentiation

Post-Synaptic Long-Term Plasticity

After a minimum of 30 minutes single pulse stimulation with responses not varying more than 10%, a 1x theta-burst induction stimulus was delivered. Single-pulse stimulation continued for 2-hours after the induction stimulus. The change in fEPSP or population spike size is then measured

Results

Acute Adult Microbiome Perturbation Leaves E-S Coupling Unchanged

Microbiome perturbation did not change the integration of synaptic excitation to generate neuronal output. $F(2, 46) = 0.312, p = 0.733$ - General Linear model

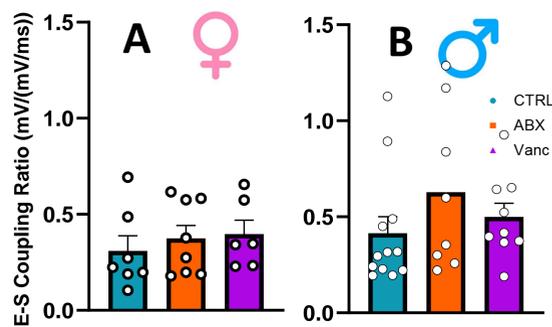


Figure 6. Bar Chart of EPSP-Spike Coupling 2 Hours post-LTP induction calculated as a ratio of Pop. Spike amplitude to fEPSP slopes for ABX, Vanc, and CTRL mice. (A) Female groups (B) Male Groups. Error bars represent SEM.

Conclusions

Microbiome perturbation through vancomycin administration may have subtle effects on the hippocampal synaptic properties of adult male mice affecting basal synaptic excitability and short-term facilitation, a process associated with working memory.

However, synaptic properties seem to prove robust to a more widescale knockdown of the microbiome through antibiotic cocktail administration in adulthood.

Results

Microbiome Depletion in Male Mice May Trend Toward Increase in Basal Synaptic Excitability

Vancomycin-treated male mice trend towards showing increased basal synaptic efficacy. Effect of treatment $F(2, 46) = 1.608, p = 0.604$ General Linear Model

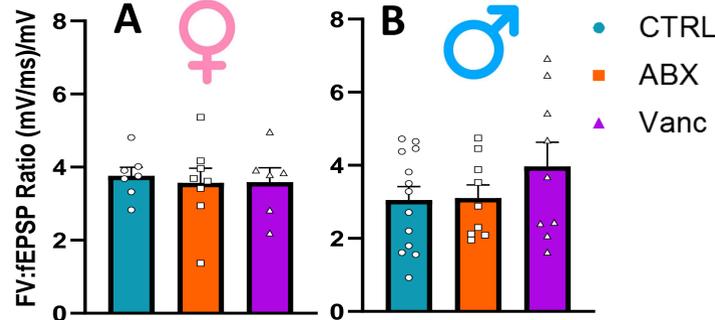


Figure 3. Ratio of evoked fibre volley amplitude to dendritic fEPSP slope is a measure of synaptic efficacy in male and female mice. (A) Female groups (B) Male Groups. Error bars represent SEM

Microbiome Depletion in Male Mice Trends Toward Deficits in Short-Term Facilitation

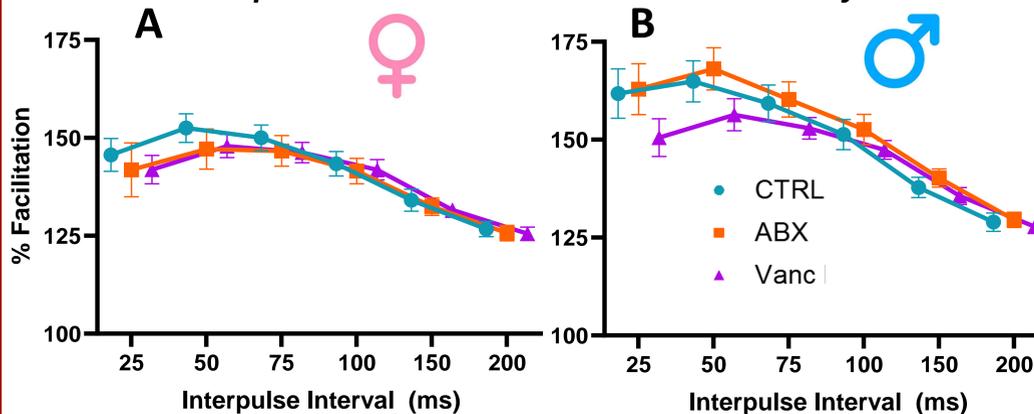


Figure 4. Percentage increase in post-synaptic response of the second of two paired stimuli at a series of inter-pulse intervals. Antibiotic-treated slices exhibited reduced short-term facilitation at 25ms and 50ms inter-pulse intervals. (A) Female groups (B) Male Groups. Error bars represent SEM.

Acute Adult Microbiome Perturbation Leaves Long-Term Potentiation Unchanged

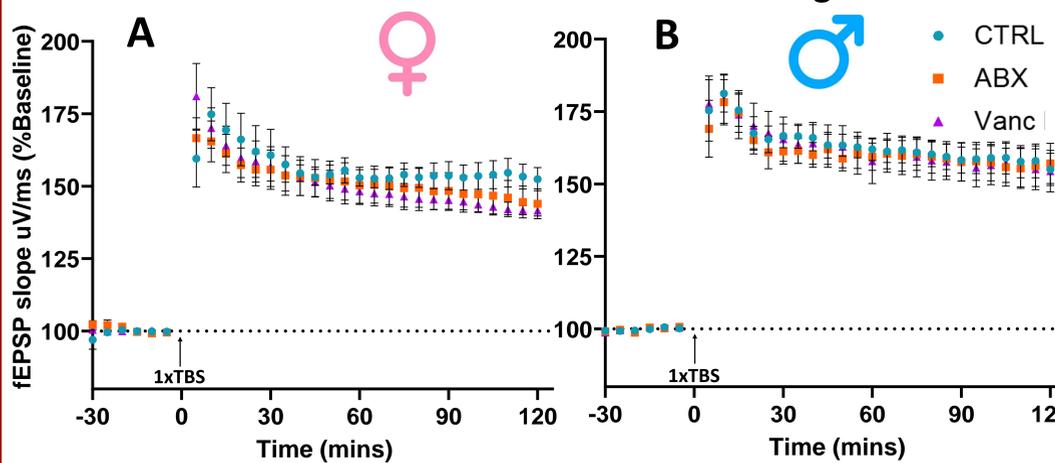


Figure 5. Timeline of CA1 dendritic fEPSP slopes for ABX, Vanc, and CTRL mice. A 30 minute baseline is followed by a potentiating 1x TBS stimulation. Responses are normalised to the average of the 30 minute baseline (A) Female groups (B) Male Groups. Error bars represent SEM

Short-term plasticity, a process implicated in working memory, trends towards impairment particularly at pulse intervals of 25ms and 50ms in vancomycin-treated male mice. Effect of treatment $F(2, 36) = 1.126, p = 0.335$ - Linear Mixed Model with Repeated Measures

Theta-burst stimulation elicits long-lasting potentiation (~150-160%) of dendritic response which was measured up to two hours. No statistically significant difference in potentiation between groups was found. $F(2, 46) = 0.574, p = 0.567$ General Linear Model

References

- Darch, Henry T., et al. "Microbial memories: Sex-dependent impact of the gut microbiome on hippocampal plasticity." *European Journal of Neuroscience* 54.4 (2021): 5235-5244.
- Gareau, Mélanie G., et al. "Bacterial infection causes stress-induced memory dysfunction in mice." *Gut* 60.3 (2011): 307-317.
- Hoban, A. E., et al. "The microbiome regulates amygdala-dependent fear recall." *Molecular psychiatry* 23.5 (2018): 1134-1144.
- Lu, Jing, et al. "Microbiota influence the development of the brain and behaviors in C57BL/6j mice." *PLoS One* 13.8 (2018): e0201829.
- Luczynski, Pauline, et al. "Adult microbiota-deficient mice have distinct dendritic morphological changes: Differential effects in the amygdala and hippocampus." *European Journal of Neuroscience* 44.9 (2016): 2654-2666.
- Luk, Berkeley, et al. "Postnatal colonization with human 'infant-type' Bifidobacterium species alters behavior of adult gnotobiotic mice." *PLoS One* 13.5 (2018): e0196510.
- Möhle, Luisa, et al. "Ly6Chi monocytes provide a link between antibiotic-induced changes in gut microbiota and adult hippocampal neurogenesis." *Cell reports* 15.9 (2016): 1945-1956.
- Pan, Jun-Xi, et al. "Absence of gut microbiota during early life affects anxiety-like behaviors and monoamine neurotransmitters system in the hippocampal of mice." *Journal of the Neurological Sciences* 400 (2019): 160-168.
- Papouin, Thomas, and Philip G. Haydon. "Obtaining acute brain slices." *Bio-protocol* 8.2 (2018).
- Further major source materials are listed here:
Cryan et al., 2019. The Microbiota-Gut-Brain axis. *Physiol Rev* 99: 1877-2013
Spichak et al., 2018. Without a bugs life. *Drug Discovery Today: Disease Models* 28: 79-93
Jaggar et al., 2020. You've got male: Sex and the microbiota-gut-brain axis across the lifespan. *Front. Neuroendocrinology*. 56: 100815