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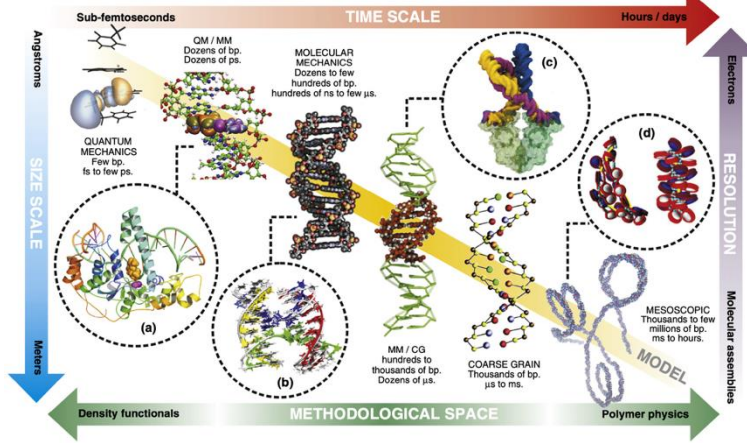
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## Computational Models of DNA

The most detailed models on the left

- calculate forces from **first principles**
- are **very accurate** and **board in scope**
- can **treat only very small systems** on very short times



The least detailed models on the right

- **omit most**, if not all **structural details**
- are more **limited and specific in scope**
- Can treat **very large systems** on very long times

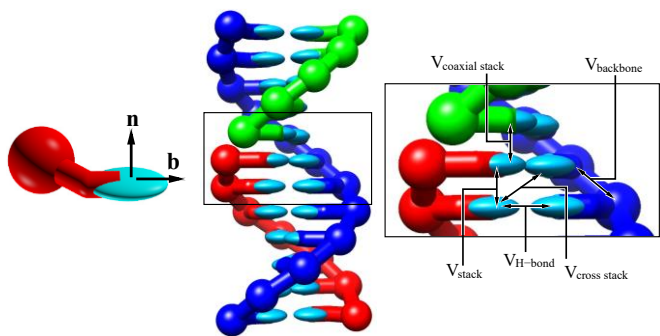
Intermediate models at the centre, which we develop, **strike a balance** as they

- **retain crucial structural details**, but **omit other less important ones** for specific purposes
- can be **quite accurate**, but have **usually a reduced scope**
- are **suitable** for modelling **medium sized systems** and **intermediate time scales**

## Coarse-Grained oxDNA Model

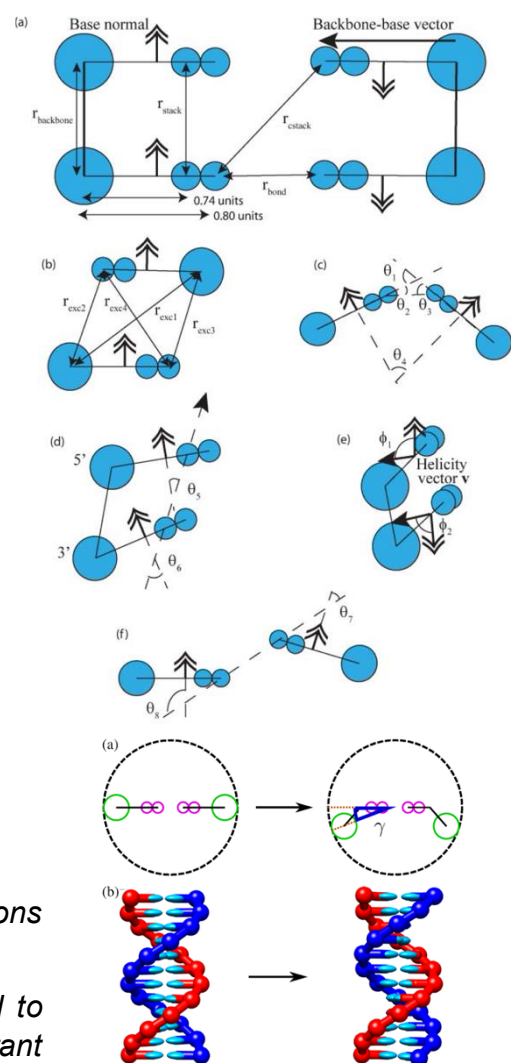
Our **oxNA suite** of DNA [1], RNA [2] and DNA:RNA hybrid [3] models is based upon what is referred to as **rigid base model**. Each nucleotide is modelled as a **rigid body** with **three interaction sites** for the effective interactions between the nucleotides, which we derive from much more detailed models. These are:

- **Connectivity** of the sugar-phosphate backbone
- **Excluded volume** of the nucleotides
- **Stacking, cross-stacking** and **coaxial stacking**
- **Hydrogen-bonding** between **complementary bases**
- **Electrostatic interaction** between negatively charged sugar-phosphate backbones



**Top and top right:** Overview of bonded and pair interactions in the oxDNA coarse-grained model of DNA.

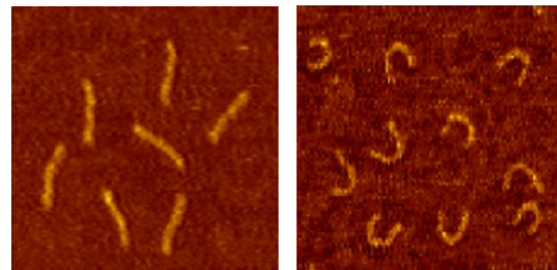
**Right:** Backbone interaction site at an angle (right) lead to the formation of the major and minor groove, an important structural feature of DNA.



## Intrinsic Curvature and Elasticity

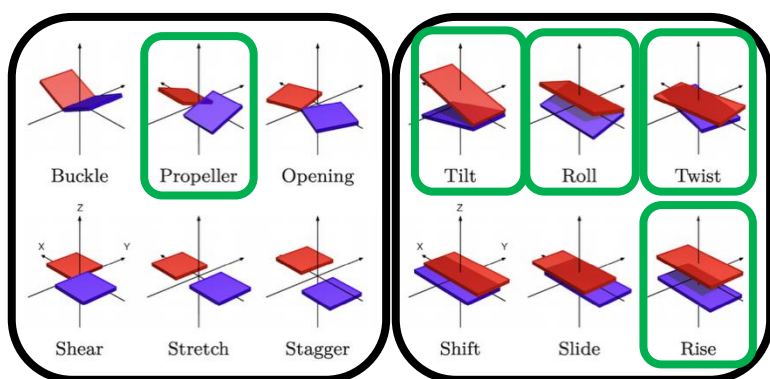
Nucleic acids are **intrinsically curved** depending on the **local sequence**. The sequence modulates also the **local elastic properties**, which is important for the **interaction of DNA or RNA with proteins**.

The sequence-dependent curvature and elasticity originate from the **relative orientation** of the nucleobases, here below displayed as slabs.



**Left:** 220 base pair long sequence of 6 base pair repeats in human telomer; **Right:** 220 base pair long sequence in *Crithidia fasciculata* (parasitic protozoa)

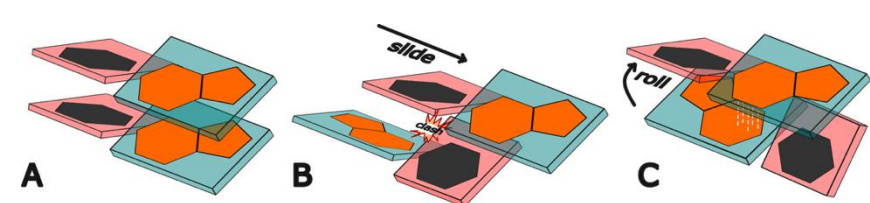
- **Twist**
  - Influenced by backbone length and stacking distance
  - +29° ... +41°
- **Roll** (soft mode)
  - Emerges due to desire of nucleobases to overlap or to avoid a steric clash
  - -2° ... +9°
- **Tilt** (stiff mode)
  - Influenced by base geometry and stacking
  - Smaller than roll -2.5° ... +2.5°
- **Rise**
  - Influenced by stacking distance
  - 3.1 ... 3.5 Å



Intra-base pair coordinates

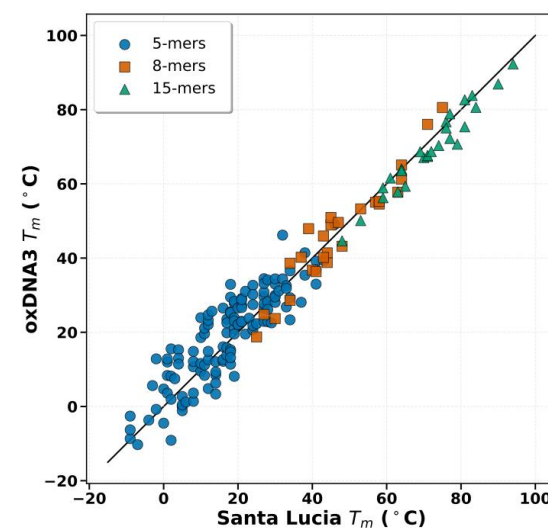
Inter-base pair coordinates

**Left:** The different size of purine and pyrimidine bases causes to either **slide (B)** or **roll (C)** within a base pair step. This effect is **physically modelled in oxDNA3**.



## oxDNA3 Results and Features

### Thermodynamics



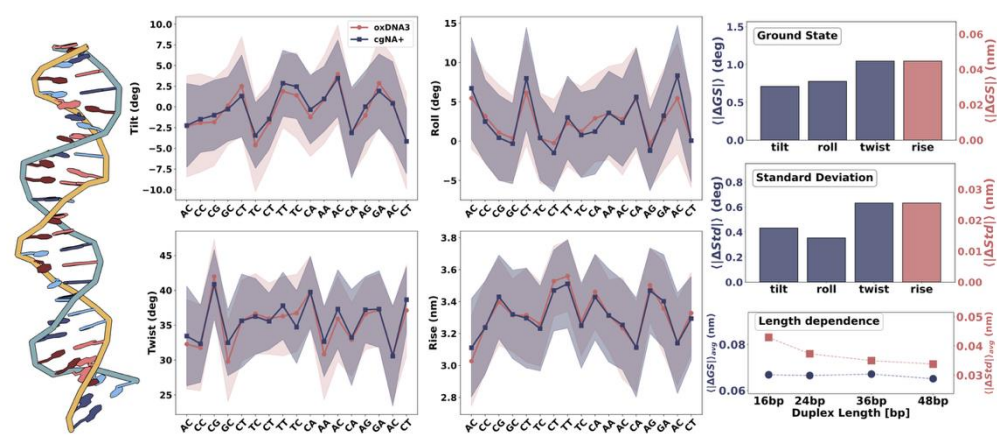
**Left:** Melting temperature in oxDNA3 compared to predictions of the Santa Lucia nearest-neighbour model

Shown are results for 129 5-mer, 30 8-mer and 30 15-mers De Bruijn sequences, which include all tetramers in the training.

The **RMSE** between oxDNA3 and Santa Lucia melting temperatures is in the region of **2.9K**.

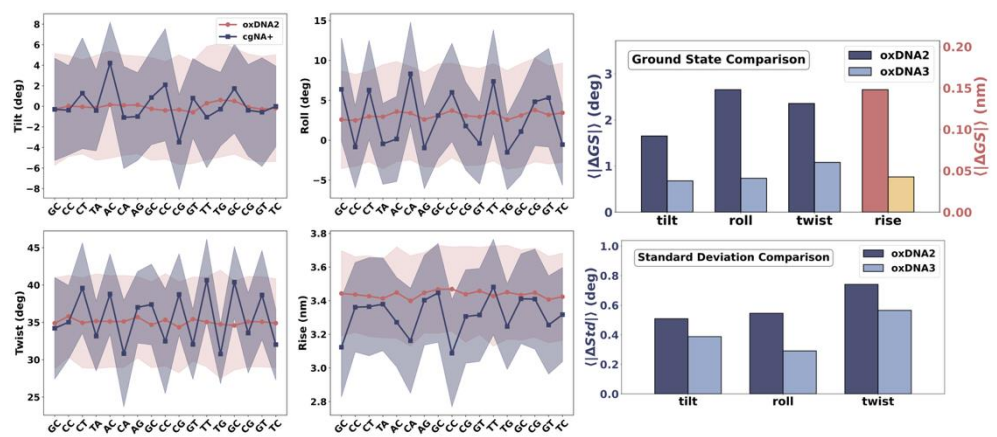
### Geometry

The geometry (**curvature** and **elasticity**) is trained on the **cgDNA+** model [4], which is itself is trained on **all-atom MD simulations** with the **parmbsc1** force field.



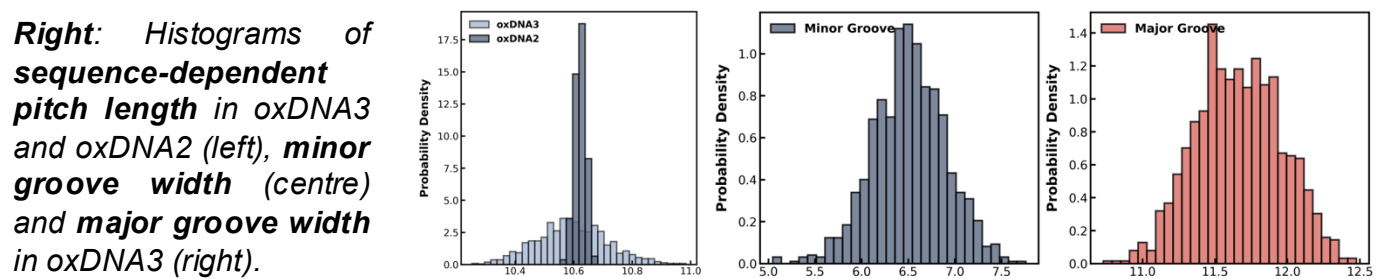
**oxDNA3 vs cgDNA+**

**Left:** Mean values of the helical coordinates **tilt, roll, twist and rise** (centre) of a random sequence (left) as well as variances (right).



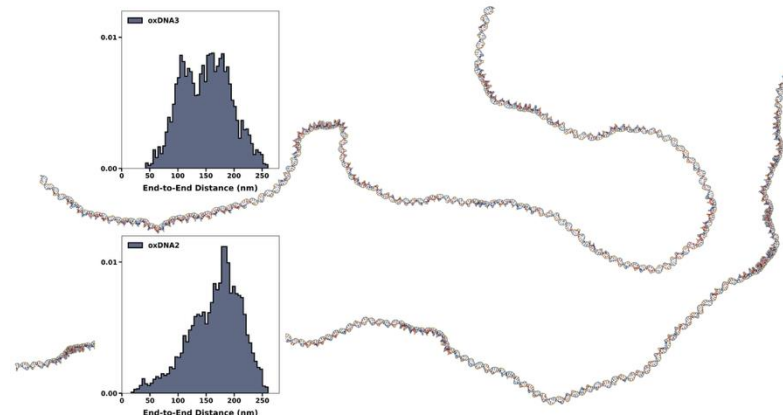
**oxDNA2 vs cgDNA+**

**Left:** Mean values of the helical coordinates **tilt, roll, twist and rise** (left) of the same random sequence and variances (right).



**Right:** Histograms of **sequence-dependent pitch length** in oxDNA3 and oxDNA2 (left), **minor groove width** (centre) and **major groove width** in oxDNA3 (right).

**oxDNA3 model (top) with sequence-specific curvature and elasticity and oxDNA2 (bottom)**



**Left:** Snapshots and distribution of the end-to-end distance of a 1,000 bp sequence from *Crithidia fasciculata*.

**Sequence-averaged persistence lengths**

**oxDNA3:**  $l_b = 45$  nm,  $l_t = 204$  nm

**oxDNA2:**  $l_b = 44$  nm,  $l_t = 163$  nm

## Machine Learning

- **Thermodynamics**
  - Cost function mean squared distance from **Santa Lucia nearest-neighbour model predicted melting temperature**

- **Geometry**
  - Cost function **Kullback-Leibler divergence** of helical coordinate distributions in oxDNA3 and cgDNA+ (parmbsc1)

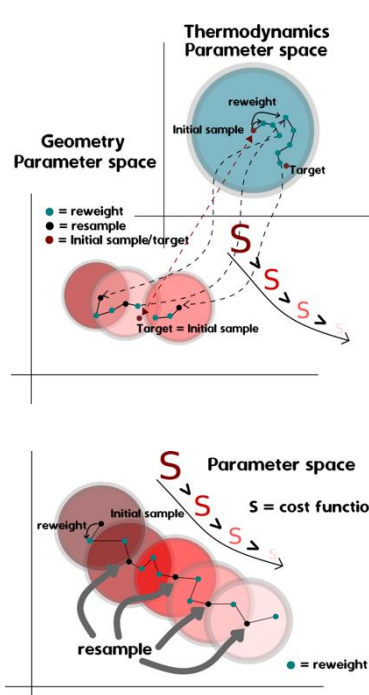
$$S[\rho_{ox}^r, \rho_{cgna}] = \int \rho_{ox}^r(\xi) \log \left( \frac{\rho_{ox}^r(\xi)}{\rho_{cgna}(\xi)} \right) d\xi$$

$\rho_{ox}^r(\xi)$  Distribution of coordinates in oxDNA

$\rho_{cgna}(\xi)$  Distribution of coordinates in cgDNA+

- **O(3000)** tuneable parameters depending on nucleotide tetramers: AAAA, AAAC, AAAG, ..., ACGT, ..., TTTT

⇒ ML framework with PyTorch using L-BFGS quasi-Newton method



## References & Acknowledgements

- [1] T. Ouldridge, A. Louis, J. Doye, *J. Chem. Phys.* **134**, 085101 (2011); B. Snodin, F. Randisi, M. Mosayebi, et al., *J. Chem. Phys.* **142**, 234901 (2015).  
 [2] P. Šulc, F. Romano, T. Ouldridge, J. Doye, A. Louis, *J. Chem. Phys.* **140**, 235102 (2014).  
 [3] E. Ratajczyk, P. Šulc, A. Turberfield, J. Doye, A. Louis, *J. Chem. Phys.* **160**, 115101 (2024).  
 [4] R. Sharma, A. Patelli, L. De Bruin, J. Maddocks, *J. Mol. Biol.* **14**, 167978 (2023).