



Preservation of Functionality, Immunophenotype and Recovery of HIV RNA from PBMC's Cryopreserved for >20 years

### John Zaunders, Wayne B Dyer, Kazuo Suzuki, Angelique Levert, Mitchell Starr and Andrew Lloyd,

St Vincent's Hospital, Sydney Sydney Blood Bank (LifeBlood), Immunovirology Research Network (IVRN).







# Speaker disclosures

- Kazuo Suzuki is a named inventor on a patent for the Double R assay for intracellular HIV RNA transcripts
- John Zaunders is a named inventor on a patent for the OX40 assay for antigen-specific CD4 and CD8 T cells

### PBMC Biobanking at Centre for Applied Medical Research, SVH, Sydney

• Major commitment for NSW State Reference Lab for HIV, at AMR

- Senior Scientist and 4 full-time staff for trials and biobanking
- Twenty -80°C Freezers
- Four Vapour Phase Liquid Nitrogen tanks

- Sydney Blood Bank HIV Research PBMC's

### Sydney Blood Bank Cohort (SBBC) – attenuated *nef*-deleted HIV-1

- 1992 5 asymptomatic HIV+ transfusion recipients
- single HIV+ donor, D36, who was also a long-term non-progressor
- normal CD4 T cell counts, after 8-10 years of infection
- Sequenced virus from the SBBC had nef / 3'-LTR deleted HIV-1

- 1995/6 Studied SBBC CD4 and CD8 T cells using fresh blood samples:
- Four colour flow: CD45RA/RO, CD28- and activated CD38+/HLA-DR+
- Compared with age-matched and transfusion-matched HIV <u>neg</u> controls
- Also, compared with other <u>viraemic</u>, <u>untreated</u> HIV+ subjects



### Original Fresh Blood CD8 T cell subsets from 1995/6





CONTROL HIV NEGATIVE COHORT

- SBBC (undetectable plasma HIV RNA)
- SBBC (detectable plasma HIV RNA)



PBMC cryopreserved in liquid N<sub>2</sub> from 3 Cohorts

- 1995 <u>Old Controls</u> HIV-uninfected from Blood Bank Study n = 20
  (mean age 60)
- 1995 <u>Old HIV+</u> Untreated, viraemic transfusion-acquired and sexuallyacquired HIV-infected subjects n = 12 (mean age 47)
- 2020 <u>Recent PBMC</u> Anonymous healthy staff donors from 2020 n = 20
   (mean age 43) IVRN QAP for PBMC storage

- Thawed, treated with DNase 1 (0.1mg/ml) 15 min RT, ~90% viability

- Stained with two 18-colour panels (PBMC subsets and T cell subsets)
- analysis on a 5-laser Fortessa
- T cell subsets in fresh blood samples from staff controls n = 16 (mean age 46)

### **PBMC** subsets – Monocyte subsets







### 18-colour PBMC subsets – B cells, Basophils, mDC and pDC





### **CD4 T cell subsets**



### **CD8 T cell subsets**



#### **CD8 T cell subsets – MAIT cells and activated CD8 T cells**



### Original Fresh Blood CD8 T cell subsets from 1995/6





CONTROL HIV NEGATIVE COHORT

- SBBC (undetectable plasma HIV RNA)
- SBBC (detectable plasma HIV RNA)



TA-LTNP COHORT

#### CD4 and CD8 T cell subsets – Thawed OLD HIV+ cells vs original fresh blood

CD45RO+ CD4 T cells





CD25+ CD134+ (OX40) Activation Induced Marker (AIM) assay of mitogen- and antigen-specific CD4 T cells

> Gated on CD3+ CD4+ Lymphocytes

No stimulation background (negative control)



Polyclonal and antigen specific CD4 T cell responses OX40 AIM (activation induced marker) 48 hour assay

Anti-CD3/CD28/CD2 OX40 AIM+ CD4



Day 7 proliferation assay of mitogen- and antigen-specific CD4 T cells

Gated on CD3+ CD4+ Lymphocytes

No stimulation background (negative control)



**Forward Scatter** 

### Optimal polyclonal T cell stimulation of PBMC via TCR in vitro

Day 4



### **Polyclonal TCR cell stimulation** of isolated CD4 T cells

Patient's

ACD

Blood

Isolated CD4 cells 96% purity



**Side Scatter** 

CD4-APC

Q3

105



Cultured for 3 days + IL2 + Anti-CD3/CD28/CD2



Polyclonal and antigen specific CD4 T cell responses 7 day proliferation assay





# Cytokine production in 48hr supernatants from Old HIV Uninfected Controls' PBMC cultured with Flu or with anti-CD3/CD28/CD2



Polyclonal and antigen specific CD4 T cell responses 7 day proliferation assay



Extract RNA to measure cell-associated HIV RNA

### **Double-R assay of HIV-1 DNA and HIV-1 RNA transcripts**



### Suzuki et al J.AIDS HIV Treatment 2019

### πCode assay >27x more sensitive than Real-Time PCR analysis

## Old HIV+ donor PBMC optimally stimulated in vitro with anti-CD3/CD28/CD2

**RNA extracted after 7 days** 

Transcripts vs donor's original plasma viral load



#### **Recovery of HIV RNA after > 20 yr storage**

Nested PCR of extracted RNA from cultured CD4 at day 7



4kbp product

#### Recovery of HIV RNA after > 20 yr storage: 4kbp product and sequencing



1758 1759 1760 1761 1762 1763 1764 1765 1766 1767 1768 1769 NC

#### Oxford Nanopore sequencing of 4kbp product

Sample ID	CA	PR	RT	IN	Drug resistance interpretation
N64 (ID.1758)	none	none	M184V	none	emtricitabine (FTC), lamivudine (3TC)
N65 (ID.1759)	none	none	D67E,T69ins,T215)	none	zidovudine (AZT), lamivudine (3TC)
N66 (ID.1761)	none	none	none	none	none
N67 (ID.1764)	none	none	none	none	none
N68 (ID.1768)	none	none	none	none	none

CA: Capsid, PR: Protease, RT: Reverse transcriptase, IN: Integrasae

### **Conclusions**

- PBMC's stored for over 20 years still contained all well defined immune subsets of cells:
  - Naïve and memory B cells and plasmablasts
  - Basophils, DC subsets, NK cell and monocyte subsets
  - CD4 and CD8 naïve and memory subsets
- Unique opportunity to compare to original flow analysis when fresh:
  - Naïve and memory and CD28- cells correlated very well
  - Activated CD4 & CD8 cells in HIV+ patient samples did not survive well
- Stored PBMC'S showed polyclonal and recall antigen-specific functions:
  - OX40 AIM assays
  - Proliferation assays
  - Cytokine assays
- After 7 days of optimal stimulation for HIV + patient PBMC samples:
  - Production of HIV RNA transcripts from latent infection
  - Enough RNA was produced for long PCR products for sequencing.

### **Future Studies**

- PBMC's stored for over 20 years are important:
  - Since late 1990's, most HIV+ patients have been on ART
  - Slow progressors, long-term non-progressors or Elite Controllers
  - Especially Elite Controllers
    - have undetectable plasma viremia without ART
    - Regarded as benchmark for functional cure
- Elite Controllers have undetectable cell-associated HIV RNA in PBMC ex vivo
  - Need multiple cultures of stimulated CD4 T cells to detect any CA-HIV-RNA



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### PBMC subsets – B cells, Basophils, mDC and pDC





### **PBMC** subsets – CD8 T cell subsets

