

#### Chlamydia And Saliva



#### **CAS Summary**

Dr Tiffany Phillips Research Fellow Melbourne Sexual Health Centre







### Study Aims

#### Background

- Increase in oropharyngeal chlamydia among MSM in Australia<sup>1</sup>
  - 1.5% prevalence among MSM attending MSHC 2015-2016<sup>2</sup>
- Saliva may play a role in gonorrhoea transmission<sup>3</sup>
  - *N. gonorrhoeae* can be isolated from saliva<sup>4</sup>
  - Saliva use as lubricant for anal sex is a strong risk factor for anorectal gonorrhoea<sup>5</sup> and tongue kissing is risk factor for oropharyngeal gonorrhoea<sup>6</sup>
- However, limited evidence for risk factors for oropharyngeal chlamydia







### Study Aims

#### **Primary Aim**

Can CT be detected in saliva in untreated MSM diagnosed with oropharyngeal chlamydia

#### **Secondary Aim**

Quantify the bacterial load of CT at the tonsillar fossae, the posterior oropharynx and in saliva







#### **Study population**

- MSM testing positive for oropharyngeal chlamydia at MSHC between August 2017 and August 2018 recruited
- Eligibility criteria:
  - No antibiotics in the previous 4 weeks
  - Returned for treatment within 14 days

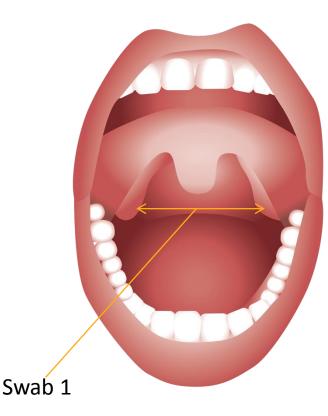






#### **Sample Collection**

- Two throat swabs:
  - First from tonsillar fossae



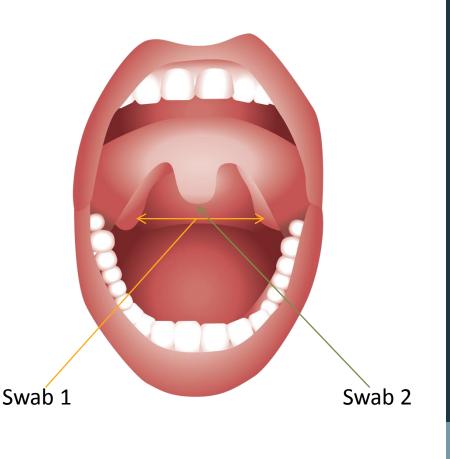






#### **Sample Collection**

- Two throat swabs:
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  - Second from posterior oropharynx



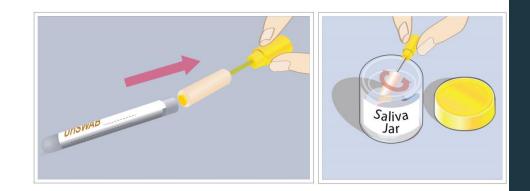






#### **Sample Collection**

- Saliva sample:
  - Participants asked to accumulate saliva for 30 seconds then expectorate into a specimen jar
  - Saliva immediately collected with a uriswab



Adapted from Chow, 2017 <sup>7</sup>







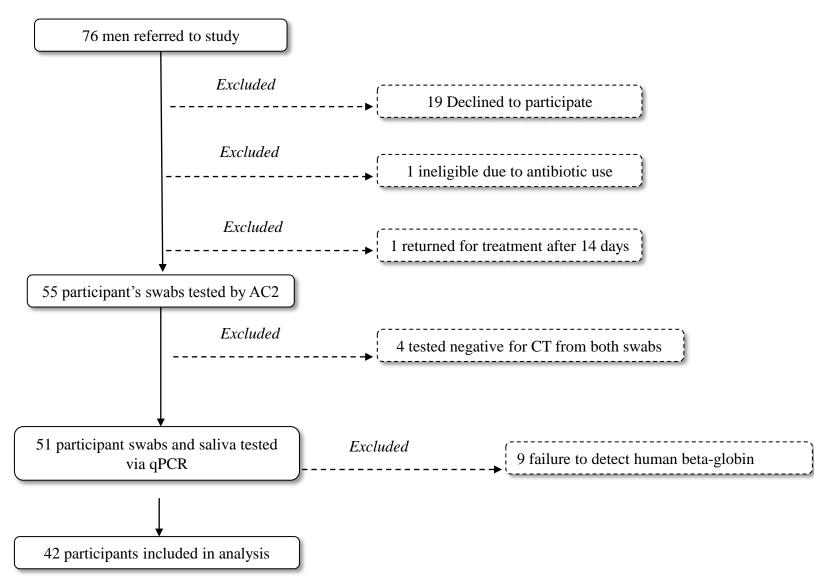
	<ul> <li>Two throat swabs collected:</li> <li>First from tonsillar fossae</li> <li>Second from posterior oropharynx</li> </ul>
Tested by qPCR –	<ul> <li>Saliva sample collected</li> <li>Participants asked to accumulate saliva for 30 seconds then expectorate into a specimen jar</li> <li>Saliva immediately collected with a uriswab</li> </ul>







#### Results



### Demographics

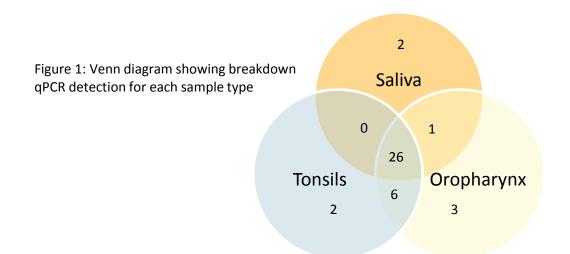
Table 1: Participant demographics and AC2 results for the 42 men included in analysis

	Age (years)	Days between diagnosis and enrolment	Sore throat	CT positive by AC2 from both tonsils and oropharynx swabs	CT positive by AC2 in tonsillar fossae swab only	CT positive by AC2 in posterior oropharynx swab only
Median [IQR]	28 [24 to 33]	5 [4 to 6]				
Number of participants (%)			12 (28.6%)	39 (92.9%)	2 (4.8%)	1 (2.4%)

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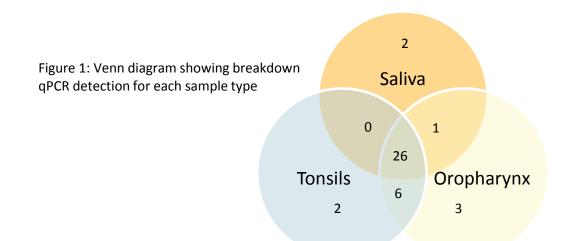


Table 2: Total median and log10 median loads of *Chlamydia trachomatis* detected by qPCR for each site

Specimen	Number of samples with CT detected from qPCR (%)	Median CT load (IQR)	Log <sub>10</sub> median CT load (IQR)
Tonsillar fossae swab	34 (81.0%)	893 copies/swab (390- 13,224)	3.0 (2.6-4.1)
Posterior oropharynx swab	36 (85.7%)	1,204 copies/swab (330- 16,211)	3.1 (2.5-4.2)
Saliva	29 (69.0%)	446 copies/ml (204-1,390)	2.6 (2.3-3.1)







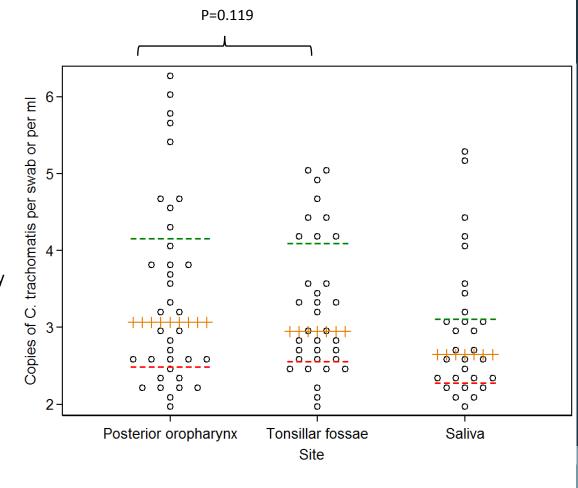


Figure 2: Log transformed median copies of *C. trachomatis* for each sample type. Amber + marks the median bacterial load for each site. Green and red dashed lines mark the upper and lower quartiles, respectively







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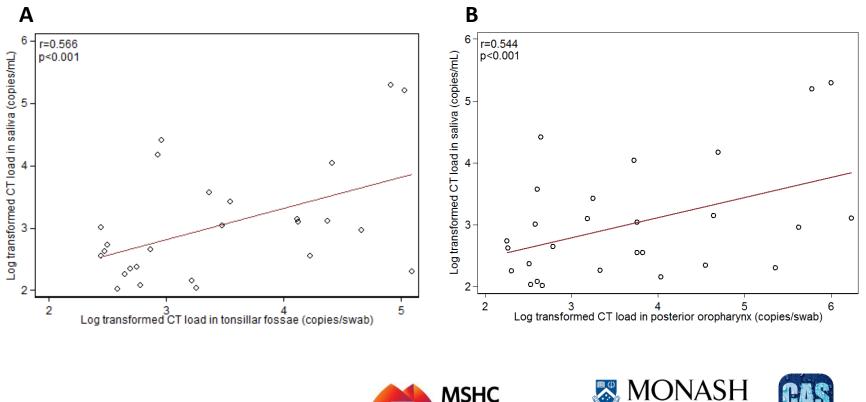
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Figure 3: Correlation of log<sub>10</sub> saliva CT load with log<sub>10</sub> CT load at the (A) tonsillar fossae and (B) posterior oropharynx



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#### Conclusions

- C. trachomatis DNA can be detected in saliva in most cases of oropharyngeal chlamydia
- There were no significant differences in the CT bacterial loads between the tonsillar fossae and the posterior oropharynx
- Saliva CT load was significantly lower than load at the tonsillar fossae and posterior oropharynx







#### Limitations

- Cannot determine viability
  - Future studies should culture CT from saliva or utilise other methods for viability detection
- Load could be influenced by the sampling methodology







### Significance

- Saliva exchange has the potential to play a role in chlamydia transmission among MSM
- The fact that three cases (7%) had only one site CT positive by AC2 suggests that sampling both the tonsillar fossae and the posterior oropharynx is necessary for accurate diagnosis of oropharyngeal CT







#### Acknowledgments

#### Study Co-Authors





Kit Fairley



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Catriona **Bradshaw** 



Marcus

Chen







Eric Chow





MELBOURNE

Jennifer Danielewski





Gerald

**Murray** 



Suzanne Garland



Jane Hocking



Fabian Kong







Vesna

De Petra

Williamson





Benjamin Howden

Rebecca Wigan

Anthony Snow



#### References

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#### Sex practices and CT load

Table 3: Median log<sub>10</sub> bacterial loads of *Chlamydia trachomatis* from qPCR stratified by those who engaged in particular sexual practice in the 30 days prior to screening and those who did not.

	Did not engage in last 30 days <sup>α</sup>	Engaged in the last 30 days	p value*
	median (IQR)	median (IQR)	
Kissing	n=4 <sup>β</sup>	n=37	
Tonsillar fossae	2.5 (2.5 to 2.5)	2.9 (2.2 to 3.6)	0.777
Posterior oropharynx	2.3 (2.2 to 3.0)	2.9 (2.3 to 4.0)	0.676
Saliva	2.6 (1.3 to 2.7)	2.3 (0 to 3.1)	0.868
Receptive penile-oral sex <sup>+</sup>	n=5 <sup>β</sup>	n=36	
Tonsillar fossae	2.6 (2.5 to 2.9)	2.8 (2.1 to 3.5)	0.629
Posterior oropharynx	2.8 (2.3 to 3.8)	2.8 (2.2 to 3.9)	0.613
Saliva	2.6 (2.6 to 2.7)	2.3 (0 to 3.1)	0.908
Receptive penile-oral sex with ejaculation <sup>†</sup>	n=26	n=15	
Tonsillar fossae	2.6 (1.9 to 3.6)	2.9 (2.6 to 3.5)	0.268
Posterior oropharynx	2.7 (2.1 to 4.3)	2.9 (2.5 to 3.7)	0.800
Saliva	2.5 (0 to 3.1)	2.3 (2.0 to 2.6)	0.896
Insertive rimming <sup>‡</sup>	n=28	n=13	
Tonsillar fossae	2.7 (2.1 to 3.4)	3.3 (2.6 to 4.4)	0.193
Posterior oropharynx	2.7 (2.2 to 3.8)	3.6 (2.5 to 4.6)	0.647
Saliva	2.3 (0 to 3.0)	2.5 (2.0 to 3.1)	0.586

#### Acknowledgments

 Mark Chung, Tiffany Rose, Sabrina Trumpour and the laboratory staff of the microbiological diagnostic unit onsite MSHC for aiding in specimen testing.